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Understanding the torrefaction of woody and agricultural biomasses through their extracted macromolecular components. Part 1: Experimental thermogravimetric solid mass loss

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A B S T R A C T

The behavior of biomass in torrefaction is determined by that of its macromolecular components, as well as by the biomass type. However, up to now, commercial compounds were typically used for modelling biomass torrefaction. This work proposes to assess the behavior of cellulose, hemicelluloses and lignin in torrefaction through extracted fractions directly isolated from woody and agricultural biomasses (ash-wood, beech, miscanthus, pine and wheat straw) thanks to an optimized extraction procedure. The solid kinetics of these extracted fractions were analyzed through thermogravimetric analysis (TGA) in chemical regime conditions (200–300 °C at 3 °C·min⁻¹ followed by 30 min at 300 °C). These experiments highlighted the influence of the biomass type and the sugar composition in the degradation of the polysaccharide fractions in torrefaction, particularly for hemicelluloses. Furthermore, the degree of preservation of the native structure of the macromolecular components, when extracting them from biomass, seems also having an impact their behavior, especially for cellulose. The comparison of the torrefaction solid kinetic profiles of these extracted fractions, dependent on the biomass type, to that of commercial compounds from previous studies suggest that these extracted fractions would be more suitable for biomass torrefaction modelling.

Keywords:

Biomass
Torrefaction
Solid mass loss
Cellulose
Hemicelluloses
Lignin

1. Introduction

The use of renewable energies and, more precisely, of biomass and waste was identified as crucial for the reduction of emissions from the European Union (EU) in the horizon 2030 [1]. Thus, biomass is already the main source of bioenergy in the EU, representing 60% of the total energy consumption from renewable sources. Forest biomass represented more than 60% for domestic biomass supplied for energy purposes in 2016, which represents around 80 Mtoe (Megatons of oil equivalent). However, the use of agricultural residues and biowaste significantly increased in the last decade [2]. This context, coupled to the high availability of biomass resources and particularly of biomass waste, requires to improve the knowhow and to optimize biomass transformation technologies.

Abbreviations: A, ash (from biomass macromolecular composition); ASE, Accelerated Solvent Extractor; C^I, «cellulose I» extracted fraction; C^{II}, «cellulose II» extracted fraction; DMSO, dimethyl sulfoxide; DMSOr, «DMSO residue» extracted fraction; E, extractives (from biomass macromolecular composition); EMAL, enzymatic mild acidolysis for lignin extraction; G, guaiacyl-lignin unit; H, hydroxyphenyl-lignin unit; H₁, «hemicelluloses 1» extracted fraction; H₂, «hemicelluloses 2» extracted fraction; L, «lignin» extracted fraction; S-, syringyl-lignin unit; TGA, thermogravimetric analyzer; XRD, X-ray diffraction; wmf, weight-moisture-free basis.

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Torrefaction is a mild thermal treatment consisting in heating biomass under an inert atmosphere, typically between 200 and 300 °C and at atmospheric pressure, for a residence time of several tens of minutes to 1 h [3]. This process is suitable for dry biomass conversion (moisture content below 60% at harvesting) [4]. Torrefaction products are composed of a torrefied solid (about 70% in mass) and some gaseous species released (about 30%) [5,6]. The torrefied solid properties are improved compared to those of raw biomass and tend to be similar to those of coal, including higher carbon content, heating value and hydrophobicity. Gaseous species released include permanent gases, mainly CO₂ and CO, and condensable species, mainly water, acids, alcohols, phenols, ketones and furans [7–9]. Temperature is the most influencing operating condition in torrefaction, and thus the yield of torrefied product decreases with it [10–12].

The macromolecular components of biomass, whose proportion and distribution in the raw material depends on the biomass type, strongly influence the transformation through torrefaction [13,14]. Previous studies demonstrated that the thermal behavior of the three macromolecular components of biomass was significantly different [15–18]. Cellulose thermal decomposition, which was described through the commercial microcrystalline cellulose, was reported to occur from temperatures close to 300 until 400 °C, the major decomposition happening between 330 and 370 °C [13]. Hemicellulose degradation starts at low to intermediate torrefaction temperatures (225–250 °C) and mostly happens in the torrefaction temperature range, until about 325–350 °C [17,19]. The different behavior of these two sugar-based fractions is thought to be due to differences in crystallinity, presence of amorphous regions, polymerization degree and sugar composition [19–21]. Indeed, while cellulose is a glucose-based polymer [22], hemicelluloses are a combination of hexose (glucose, mannose, galactose) and pentose sugar units (xylose, arabinose) [23,24]. On the contrary, lignin is based on a polyaromatic structure, based on phenylpropane units: H- (hydroxyphenyl-), G- (guaiacyl-) and S- (syringyl-) units [25]. Lignin typically follows a progressive degradation during torrefaction, from about 200 °C [26,27] to 250 °C [28] until high pyrolysis temperatures about 500 °C [29], 600 °C [30], 720 °C [27] or even 900 °C, in function of the heating rate [3]. As a result, lignin is only partially degraded in the torrefaction temperature range.

Extractives and ash are also part of the biomass macromolecular composition. However, their influence in biomass degradation through torrefaction was still not fully elucidated. In the case of extractives, most of the species present lower volatilization temperatures suggesting a probable release below 200 °C [31,32]. The impact of ash on biomass thermal decomposition was clearly identified at pyrolysis [33,34] and gasification temperatures [35,36]. In torrefaction, a catalytic effect of potassium on mass loss during torrefaction was suggested [37–40]. However, the representativeness of inorganic elements on the raw biomass on these torrefaction experiments is controversial, as they were based either on an impregnation with commercial compounds or on a modified biomass structure through doping. When comparing raw biomasses of the same nature with a different ash composition, no correlation was clearly identified under torrefaction conditions [41].

Up to now, the behavior of biomass macromolecular components in torrefaction was usually characterized by studying commercial compounds [8,13,14,17]. The reason of this approach is the complex structural matrix of biomass [42–44], which makes the extraction of representative fractions of cellulose, hemicelluloses and lignin directly from the raw material a real challenge.

Cellulose is generally extracted from biomass through acid hydrolysis, after a complete lignin removal by a chlorite treatment.

The objective of this treatment is hemicellulose decomposition due to their higher lability compared to cellulose. However, it may also affect cellulose reactivity by degrading cellulose amorphous areas and drastically reducing the degree of polymerization of cellulose chains. A weaker acid treatment may contribute to remove only the amorphous part of the structure, while cellulose crystalline regions with allomorph structure type I from native cellulose, more resistant to hydrolysis, would be preserved [45]. The alkaline solubilization of residual hemicelluloses would be an alternative to the acid hydrolysis [46,47]. The main advantage of this treatment is the preservation of cellulose amorphous regions and of a higher degree of polymerization. However, the high alkali concentration induces changes in the cellulose allomorphic structure due to mercerization (conversion of native cellulose structure type I into a different allomorph, the so-called cellulose II) [48].

A first main part of hemicelluloses in biomass can be extracted through dimethyl sulfoxide (DMSO) [49,50]. This treatment is applied to delignified biomass, typically after a chlorite treatment for lignin removal [51]. Its main advantage is the preservation of hemicellulose acetyl groups [50]. A second hemicellulose fraction can be isolated from the residue of the DMSO extraction through a strong alkaline treatment, so as to separate cellulose and residual hemicelluloses. Nevertheless, compared to the first fraction, hemicellulose structure is slightly modified, principally in terms of its acetyl group content, removed by the alkaline treatment [52].

Lignin is generally extracted from biomass through different procedures [25,53,54]. Lignin acidolysis using dioxan seems to provide the best compromise between purity and yield. That is the reason why it is one of the most frequently used methods for lignin extraction [55]. Milled wood lignin is obtained through long ball milling followed by purification steps and a dioxan extraction. However, the obtained purity for lignin is moderate and the production yields are low. Enzymatic mild acidolysis for lignin extraction (EMAL) was recently proposed [56]. Even if the yield of production remains low, EMAL allows to obtain lignin of excellent purity. Finally, the Klason procedure was shown to be suitable to quantify lignin in biomass, but not to extract it, as its strong acid treatment has an effect on the condensation degree of the lignin structure [57].

The objective of this study is to assess the contribution of biomass main macromolecular components, namely cellulose, hemicelluloses and lignin, on their solid transformation through torrefaction. To achieve this, an optimized extraction procedure was developed to isolate extracted fractions corresponding to these biomass main macromolecular components, so as to get an improved representativeness of their behavior compared to the classically used commercial compounds. In the first part of this paper, solid degradation profiles of the cellulose, hemicelluloses and lignin fractions in torrefaction were studied versus time and temperature in a thermobalance. In the second part of this paper, a torrefaction kinetic model able to predict biomass solid mass loss in function of its macromolecular composition and biomass type is proposed, based on these experimental results.

2. Material and methods

2.1. EXTRACTION PROCEDURE

Fractions representing the main biomass macromolecular components were extracted according to a sequence of different methods available in the literature with some in-house adaptations. The acronyms of the extracted fractions are presented in the legend (Fig. 1). Subscripts and Arabic numerals (1 and 2) were used for complementary fractions of the same macromolecular component, as it was the case for hemicelluloses. Superscripts and Roman

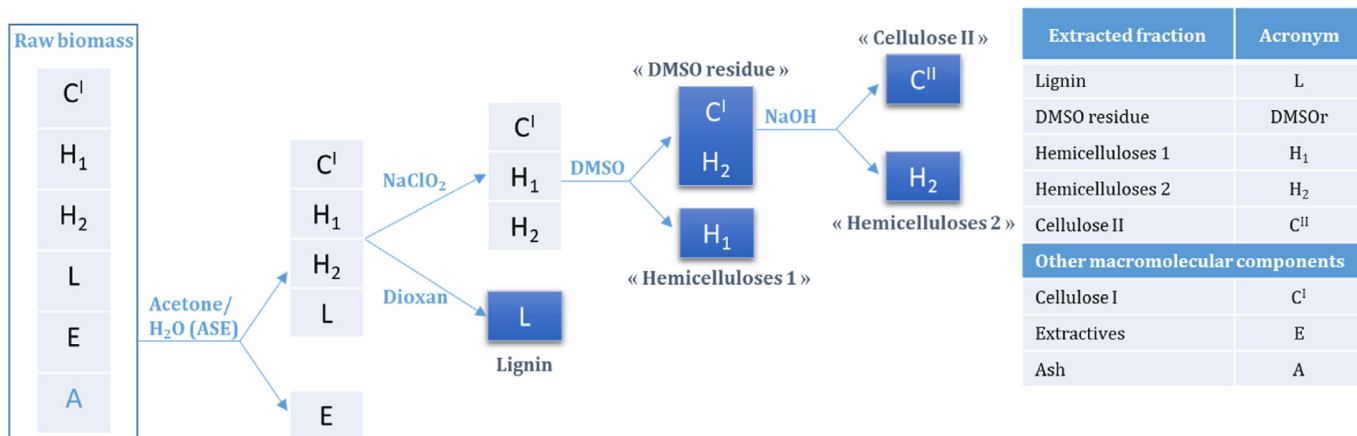


Fig. 1. Extraction procedure of the fractions representing the main biomass macromolecular components and extracted fractions obtained from raw biomass.

numerals (I and II) were indicated for different allomorphs of the same macromolecular component, namely cellulose.

Biomass, both wood chips, agricultural straw and grass, were firstly ground using a laboratory 5-knife mill (Pulverisette) equipped with 1 mm sieve.

First of all, extractives were removed using an ASE (Accelerated Solvent Extractor) with 2 cycles at 1500 psi, firstly with water at 110 °C then with acetone at 95 °C equipped with 12 independent 200 mL extraction cartridges with an automatic autosampler [58]. The extraction cells were filled up with a mixture of biomass powder (size particle below 1 mm) and glass beads. The extracted material was air-dried during one week before lignin or polysaccharides extraction.

Lignin fraction (L) was extracted by refluxing the extractives-free biomass powder in a dioxan/water (85/15 v/v) mixture containing 0.01 M HCl for 4 h under nitrogen [55]. The solubilized lignin was filtered out and precipitated by evaporating half of the volume of the solution using a rotative evaporator under vacuum. Water was added and the previous steps were repeated five times to allow a complete removal of dioxan. The precipitated lignin was then centrifuged, washed with water to neutrality, and finally freeze-dried.

The acidic medium used in lignin extraction could partially hydrolyze cellulose and hemicelluloses. Therefore, polysaccharide fractions were isolated from a new sample of extractives-free biomasses. Before separating polysaccharides, it was necessary to perform a complete delignification of the sample. Wood powder was first delignified using sodium chlorite in an acetate/acid buffer (pH = 4.6) in a 5 L glass reactor (liquor/biomass weight ratio = 6), at 60 °C, under mechanical stirring and nitrogen gas flow [59]. Sodium chlorite was progressively added to the medium until complete delignification. The measurement of Kappa number (carried out using the ISO 302–2015 standard method) of the treated biomass samples was used to follow delignification. Extraction time varied between one and two weeks depending on the lignin content and raw materials. At the end of the delignification, biomass particles were totally converted into fibers, which were thoroughly washed with demineralized water until neutral pH.

Two fractions of hemicelluloses were obtained by combining two successive treatments, an extraction through dimethyl sulfide (DMSO) and an alkaline hydrolysis.

The first fraction, called here « hemicelluloses 1» (H₁), was obtained by a mild treatment based on an extraction with DMSO in a 5 L glass reactor (liquor/wood weight ratio = 6), at room temperature, under mechanical stirring and nitrogen gas flow. After the

extraction, the liquid phase was separated by filtering over Buchner filter (porosity 2), and the solid fraction, called here « DMSO residue» (DMSOr), thoroughly washed with deionized water. Because of the high boiling point of DMSO, it was not possible to remove it by evaporation under vacuum. H₁ fraction was then obtained by dialyzing the DMSO of the solution. The liquid phase was distributed in 20 small bags of cellophane membranes (cut-off MW 3600) which were placed in a large volume of distilled water (20 L). The dialysis took place during two weeks and water was changed twice a day. Hemicelluloses were then recovered by freeze-drying and lyophilization.

The second fraction, called here « hemicelluloses 2» (H₂) was obtained by treating DMSOr fraction in alkaline conditions (NaOH, 2.5 M, 60 °C, 30 min, liquor/biomass weight ratio = 6). After the extraction, the liquid phase was separated by filtering over Buchner filter (porosity 2). H₂ fraction was also obtained through dialysis as described above but in this case the dialysis was carried out until the conductivity of the water bath remained unchanged for 24 h, indicating that all the NaOH was completely removed. Hemicelluloses were also recovered by freeze-drying and lyophilization.

The solid fraction obtained after the alkaline treatment was neutralized with diluted HCl, dialyzed with membranes and lyophilized. This fraction, called « cellulose II» (C^{II}), contains essentially cellulose from the allomorphic type II.

As H₁ and H₂ are sensible to heat, these fractions were lyophilized so as to remove water without heating, while C^{II} were directly air-dried. In the case of DMSOr, a part of it was lyophilized for the XDR crystallinity tests while a second part was dried at 105 °C for 24 h for torrefaction experiments. Lyophilization allowed to obtain polysaccharide fractions under the shape of a cotton-like material, which simplified their handling in torrefaction experiments. L fraction was dried at room temperature for several days and ground manually.

2.2. CARBOHYDRATE COMPOSITION

The carbohydrate composition was determined after a second acidic post-hydrolysis of the liquor to depolymerize all remaining oligomers to monosaccharides using diluted H₂SO₄ aqueous solution (2%) for 1 h at 121 °C in an autoclave [63]. Monosugar content was determined from ionic liquid chromatography after acidic hydrolysis of the polysaccharides. Monosugar analysis was carried out after a two-step acidic hydrolysis of wood and pulps, by the ASTM method E1758 - 01 (2007). Quantification of neutral monosugars was obtained on a DIONEX HPAE-PAD ion chromatograph equipped

with a pulsed amperometric detector.

2.3. XRD CHARACTERIZATION

X-Ray diffraction (XRD) characterization was carried out for C^{II} and DMSOr fractions for investigating possible changes in cellulose structure. For this purpose, an XPERT-PRO MPD Diffractometer system was used, with the measuring program from PANalytical. The anode material was copper, with a selected wavelength $\lambda_{\text{Cu}} = 1.5419 \text{ \AA}$. The reflection method was Bragg Brentano type.

2.4. THERMOGRAVIMETRIC ANALYSIS

Remaining solid mass of the extracted fractions through torrefaction was followed in function of time and temperature in a thermogravimetric analyzer (TGA). A three-plate crucible (10 mm diameter) allowed to use a higher mass than in typical torrefaction experiments [60]. Its suspended position in the TGA oven, as well as the maximum bed thickness of sample per plate (2 mm), were set so as to ensure chemical regime conditions for the three sample layers [61]. About 100 mg of sample were used for experiments involving L, C^{II} and DMSOr fractions. The low density of the hemicellulose fractions limited their mass to 50 mg for H₁ and to 25 mg for H₂.

Samples were torrefied in the thermobalance under a 50 mL/min helium (He) flow. The heating program started by a sample pre-heating from room temperature to around 70 °C at a heating rate of 3 °C·min⁻¹. At this point, the stabilization was achieved between the internal heating of the oven and the cooling water temperature. Then, a second pre-heating stage was carried out until 200 °C, at 3 °C·min⁻¹, which was considered as the reference

temperature for the beginning of torrefaction. At 200 °C, the moisture content can be considered as negligible as reactions associated to torrefaction have not started yet. From room temperature to 200 °C, mass loss was neglected (typically below 3%) [62]. Torrefaction experiments were then carried out by heating the sample from 200 to 300 °C at 3 °C·min⁻¹, and then by keeping it at 300 °C for 30 min.

The repeatability of torrefaction experiments was checked by carrying out each experiment twice. The standard deviation in mass loss calculations was below 0.5%. The results of the thermogravimetric analysis are expressed in weight-moisture-free basis (wmf).

3. Results and discussion

3.1. CHARACTERIZATIONS OF THE EXTRACTED FRACTIONS

3.1.1. Composition of the polysaccharide-based fractions

Polysaccharide based-fractions were characterized in terms of sugar composition and acetyl group content (Table 1) as defined in Ref. [41,58,63]. Sugar composition of the raw biomasses was also added to Table 1 for comparison [41].

Xylose composition was quite similar for the two hemicellulose fractions, except for pine fractions, for which xylose is principally present in H₂ fractions (Table 1). Mannose was mostly found in wheat straw and miscanthus H₂ fractions. However, in the case of pine, most of the galactose was found in H₁ fraction, together with mannose, as they are associated in galactomannan and galactoglucomannan sugars in coniferous wood [51]. This might suggest a different stability of the mannose-based sugars in function of the biomass type. Acetyls were mostly present in H₁ fractions, as

Table 1
Neutral monosugar distribution and functional groups of the polysaccharide fractions.

Extracted fractions	Neutral sugars balance (normalized)					Functional groups
	Glucose	Xylose	Mannose	Galactose	Arabinose	Acetyl groups
	%*					% wmf**
«Hemicelluloses 1» (H₁)						
Ash-wood	5.5	72.4	0.0	0.3	0.6	21.2
Beech	0.2	75.9	0.0	0.4	0.6	22.9
Miscanthus	12.5	64.4	0.0	0.7	5.9	16.5
Pine	14.8	18.4	49.2	4.1	3.0	10.5
Wheat straw	32.3	52.0	0.0	1.6	4.9	9.1
«DMSO residue» (DMSOr)						
Ash-wood	77.3	14.9	1.8	0.0	0.8	5.2
Beech	78.3	16.2	0.0	0.0	0.4	5.0
Miscanthus	76.6	11.9	5.9	0.2	0.3	5.2
Pine	78.7	16.2	0.0	0.0	1.2	4.0
Wheat straw	78.5	5.4	11.5	0.2	0.5	3.8
«Hemicelluloses 2» (H₂)						
Ash-wood	16.7	75.8	0.0	0.8	5.8	0.9
Beech	12.3	79.9	0.0	0.8	6.2	0.8
Miscanthus	9.8	86.4	2.0	0.6	0.6	0.7
Pine	6.2	80.2	7.5	0.7	4.2	1.2
Wheat straw	9.9	46.6	33.3	3.1	6.4	0.8
«Cellulose II» (C^{II})						
Ash-wood	98.2	0.0	1.8	0.0	0.0	n.m.
Beech	100.0	0.0	0.0	0.0	0.0	n.m.
Miscanthus	100.0	0.0	0.0	0.0	0.0	n.m.
Pine	99.1	0.0	0.9	0.0	0.0	n.m.
Wheat straw	100.0	0.0	0.0	0.0	0.0	n.m.
Raw biomasses [41]						
Ash-wood	69.2	24.9	2.0	1.1	2.7	3.6
Beech	63.9	29.1	3.0	2.0	2.0	8.3
Miscanthus	69.9	26.1	0.5	0.7	2.9	2.6
Pine	70.6	8.2	16.9	2.8	1.6	1.7
Wheat straw	63.7	29.7	0.8	1.6	4.2	1.7

*% of the total monosugars; **% wmf = weight-moisture-free; n.m.: not measured.

the alkaline treatment to obtain the second fractions destroys the acetate functional group in hemicelluloses. These differences in composition indicated that both hemicellulose fractions are independent.

DMSOr fractions were mainly constituted of cellulose, as it was mainly composed of glucose (Table 1). However, hemicellulose sugars and acetyl groups were also present in considerable amounts. The similar glucose to xylose ratios in DMSOr fractions were close for all the extracted fractions, which indicated a similar proportion of cellulose and hemicelluloses in these fractions. All DMSOr fractions, except that from wheat straw, presented higher xylose content compared to mannose content. This is also the case for all raw biomasses, except pine. As a result, a higher recovery of mannose was achieved in the H₁ fraction for all biomasses except for wheat straw.

Sugar distribution of the C^{II} fractions proved their high purity, as they were principally constituted of glucose and only minor amounts of mannose for ash-wood and pine celluloses (Table 1). Commercial microcrystalline cellulose, i.e. Avicel, has typically a glucose content of 96.1 %wmf, a xylose content of 2.4 %wmf and a mannose content of 1.5 %wmf [64], which represents a lower purity compared to C^{II} fractions.

3.1.2. Cellulose allomorphs

The final alkaline treatment to the DMSO residue in the extraction procedure lead to a high purity in glucose for the C^{II} fractions. However, the operating conditions used in this step were suspected to induce mercerization. The mercerization in alkaline medium is due to Na⁺ ions going into the cellulose structure and separating glucan chains. Then, these chains recrystallize in an anti-parallel structure with higher stability, named « cellulose II » [65], which differs from native cellulose in biomass, called « cellulose I » and formed by parallel glucan chains. X-Ray diffraction (XRD) characterization was then carried out on C^{II} and DMSOr fractions for investigating possible changes in cellulose structure (Fig. 2).

XRD results confirmed cellulose mercerization, this is, a conversion of cellulose allomorphic structure type I (Fig. 2, left) in all DMSOr fractions to cellulose II in all C^{II} fractions (Fig. 2, right). That is the reason why the final solid residue was called C^{II} in this work. During the whole extraction procedure, cellulose I was preserved in all steps before the alkaline extraction (until the DMSOr fractions). As a result, DMSOr fractions was also analyzed in terms of characterization and behavior in torrefaction. Therefore, by comparing the behavior in torrefaction of DMSOr and C^{II} fractions, the influence of cellulose allomorphic structure on its conversion in torrefaction could be assessed. Furthermore, the influence of the

presence of hemicellulose sugars on a cellulose fraction could also be evaluated by comparing the behavior in torrefaction of H₁ and H₂ fractions to that of DMSOr fraction.

3.2. TORREFACTION EXPERIMENTS

3.2.1. Cellulose fractions

Both C^{II} and DMSOr fractions were studied due to their different cellulose allomorphs (Fig. 2). Furthermore, the study of DMSOr fractions allowed to assess the influence of the presence of hemicellulose sugars in a cellulose fraction.

3.2.1.1. «Cellulose II » fraction (C^{II}). The remaining solid mass profiles were generally similar for all « cellulose II » fractions (C^{II}) for all biomass types (Fig. 3, left). The final remaining solid mass at the end of the non-isothermal torrefaction was similar for all samples, varying from 92.2 to 86.6 %wmf. Then, the final remaining solid mass for C^{II} fractions ranged between 66.0 and 55.8 %wmf at the end of the isothermal torrefaction at 300 °C.

Degradation rate curves (Fig. 3, right) showed similar profiles for ash-wood and wheat straw C^{II} fractions on the one hand, and for beech and pine C^{II} fractions on the other hand. The acceleration in the degradation of these fractions was progressive from low torrefaction temperatures (230 °C). In the isothermal step, the degradation rate of C^{II} samples started at around 1.0 %wmf·min⁻¹ for pine and 1.4 %wmf·min⁻¹ for ash-wood C^{II}. Degradation rates were rather constant versus time for pine and beech fractions, while they decreased progressively with time for the other fractions. The degradation rate at the end of the experiment was in the same order of magnitude for all samples (around 1.0 %wmf·min⁻¹).

The degradation of C^{II} fractions was observed from temperatures (around 230 °C) lower than those reported previously for commercial microcrystalline celluloses such as Avicel [17,26,27,66]. However, the reported degradation rate at 300 °C was much lower for commercial cellulose compared to C^{II} fractions [14]. This result might be due to the higher thermochemical reactivity of commercial Avicel, due to the acid hydrolysis employed for its extraction, typically carried out on a bleached sulfite pulp [66]. The milder operating conditions of the extraction procedure selected in this study would less impact cellulose reactivity, by contributing in the preservation of cellulose from native biomass. As a result, a higher degree of polymerization might be expected, as well as the preservation of the amorphous areas from native cellulose, which is not the case with the commercial acid hydrolysis. Furthermore, a higher purity in glucose was achieved for cellulose extracted fractions (Table 1), compared to reported values for commercial

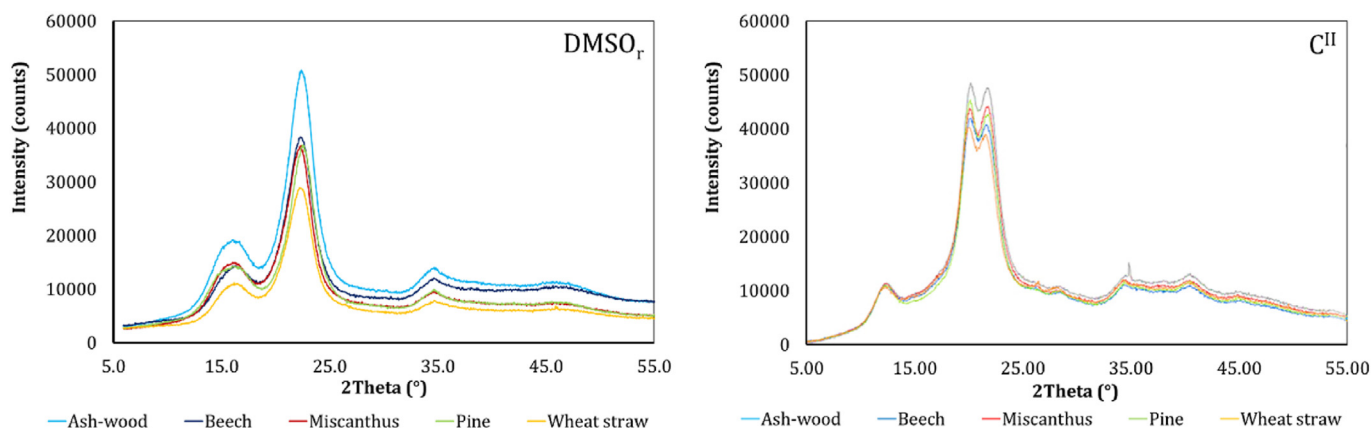


Fig. 2. XRD spectra of DMSOr (left) and C^{II} (right) extracted fractions from the 5 biomasses.

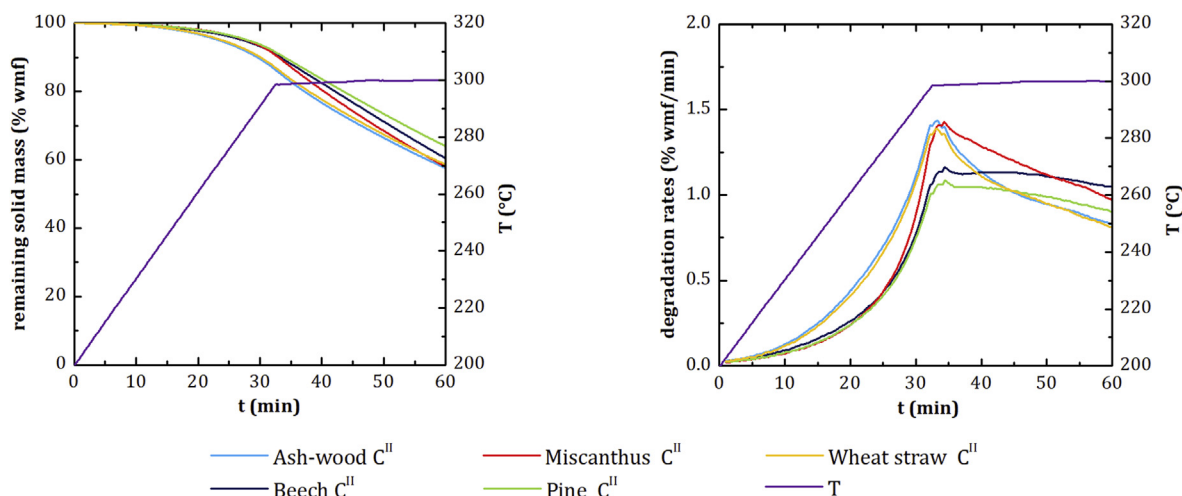


Fig. 3. Remaining solid mass (left) and degradation rates (right) versus time and temperature in torrefaction in TGA-GC/MS for the C^{II} fractions from the biomasses of study.

microcrystalline cellulose, which might also impact its reactivity.

3.2.1.2. «DMSO residue » fraction (DMSOr). «DMSO residue» (DMSOr) fractions degradation profiles and extent were shown to be dependent on the biomass type (Fig. 4). The final remaining solid mass at the end of the isothermal step was between 42.1 %wmf for ash-wood DMSOr and 64.1 %wmf for pine DMSOr (Fig. 4, left). Degradation rates were below 1.9% wmf·min⁻¹ for all DMSOr fractions (Fig. 4, right). Degradation rate profiles presented a similar shape to those of the equivalent C^{II} fractions. However, the degradation extent of this fraction in torrefaction was higher, compared to that of C^{II} fractions, presumably because of the presence of hemicellulose sugars (Table 1).

Three main solid degradation profiles could be distinguished for DMSOr fractions through the degradation rate profiles (Fig. 4, right). First of all, ash-wood DMSOr presented a degradation rate profile characterized by a global and soft mass loss acceleration. This kind of profile was also observed for beech, whose deceleration phase was shorter, from around 265 to 275 °C. The second kinetic profile corresponded to miscanthus DMSOr, whose behavior was close to that of deciduous wood, with a clear distinction between acceleration and deceleration steps and without the last acceleration step before 300 °C. Finally, pine DMSOr presented a

different solid kinetic behavior, which consisted in an acceleration of the mass loss until the end of the non-isothermal torrefaction. The degradation profile of wheat straw DMSOr was intermediate between the two previous types. It presented a slight change in the mass loss acceleration around 285 °C, which might be due to the higher mannose content, less reactive than xylose, compared to the other DMSOr fractions.

In the isothermal step, degradation rate profiles could be classified in the same groups. It is noteworthy that the final remaining solid mass tends to stabilize around 0.7–0.8% wmf·min⁻¹ whatever the biomass was, while their degradation rate values at the beginning of this step were relatively different (1.2–1.7% wmf·min⁻¹). This result might indicate that solid mass loss below 300 °C could principally correspond to that of hemicellulose sugars, due to the differences in the xylose and mannose composition of the samples. However, the degradation in the isothermal step would rather only correspond to that of glucose, whose content is similar for all DMSOr fractions. Indeed, degradation rate profiles at 300 °C for DMSOr (Fig. 4) and for C^{II} (Fig. 3) fractions were similar. This might indicate that the change in cellulose allomorphic structure due to the extraction procedure did not significantly impact cellulose solid kinetics in torrefaction.

To sum up, the presence of hemicelluloses in the DMSOr fraction

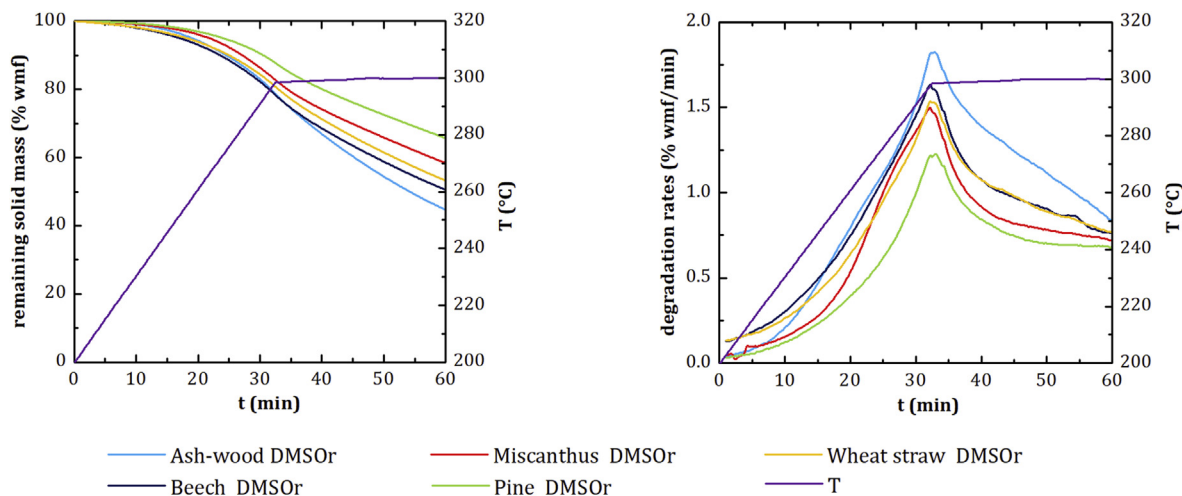


Fig. 4. Remaining solid mass (left) and degradation rates (right) versus time and temperature in torrefaction in TGA-GC/MS for the DMSOr fractions from the biomasses of study.

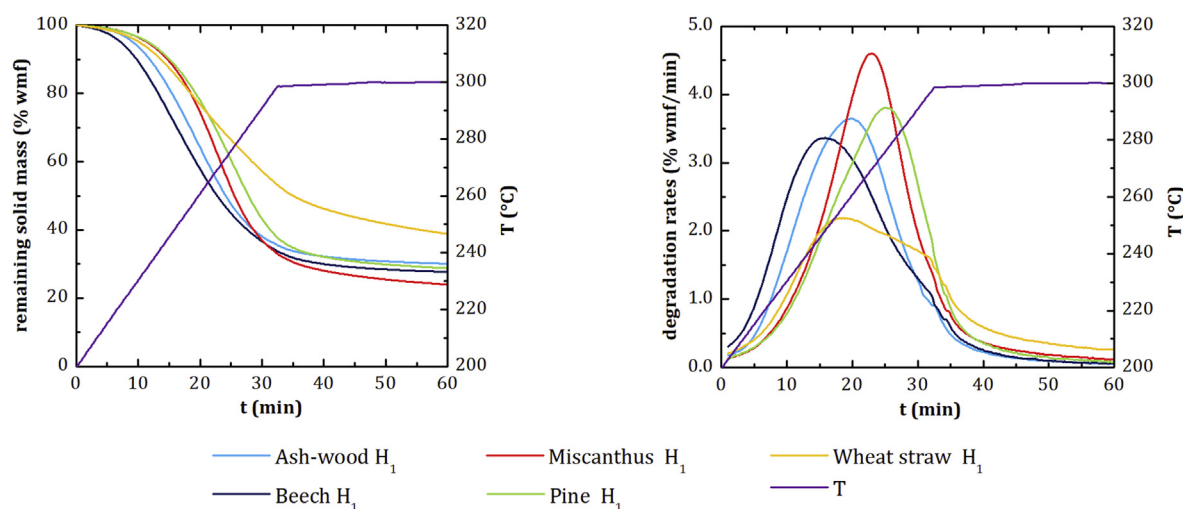


Fig. 5. Remaining solid mass (left) and degradation rates (right) versus time and temperature in torrefaction in TGA-GC/MS for the H₁ fractions from the biomasses of study.

leads to a more enhanced solid mass loss, especially below 300 °C. The change in cellulose allomorphic structure does not seem to significantly impact its degradation in torrefaction.

3.2.2. Hemicellulose fractions

3.2.2.1. «Hemicelluloses 1» fraction (H₁). The remaining mass loss at the end of the non-isothermal torrefaction was relatively close for all «hemicelluloses 1» fractions (H₁), between 32.0 and 36.7%wmf, except for wheat straw (52.1%wmf, Fig. 5, left). This value moved to around 38.3%wmf for wheat straw H₁ fraction and 23.7%wmf for miscanthus H₁ fraction at the end of the isothermal torrefaction step. These results are in agreement with previous studies pointing out the high extent of hemicelluloses degradation through torrefaction [8,17,18,67]. Maximum degradation rates were between 2.0% wmf·min⁻¹ for wheat straw H₁ fraction and 4.7% wmf·min⁻¹ for miscanthus H₁ fraction (Fig. 5, right).

Solid degradation profiles were to be similar for all H₁ samples, except for wheat straw H₁ fraction (Fig. 5, right). It is noteworthy that the maximum degradation rate was not reached at the same temperature for all H₁ fractions (Fig. 5, left).

Similarities in solid kinetics during the non-isothermal torrefaction step allowed to classify H₁ fractions in three groups. First of all, similarities were found in the behavior of H₁ fractions from deciduous wood (from ash-wood and beech), which may be derived from their similar composition in xylose (around 75% wmf) and acetyl groups (around 22% wmf). Ash-wood and beech H₁ fractions were characterized by a regular acceleration of the solid mass loss from about 210 °C. This acceleration seemed to start a little later for ash-wood H₁ fraction, compared to the corresponding fraction from beech. The maximum degradation rate was achieved at 245 °C for beech and slightly later for ash-wood H₁ fraction, around 260 °C.

Miscanthus H₁ fraction presented a very close behavior to that of the previous H₁ fractions. However, the acceleration of the solid mass loss occurred at slightly higher temperatures (around 220 °C). Furthermore, it showed a steeper slope until the highest degradation rates, which were reached at around 270 °C. These differences with deciduous wood might be explained by the higher glucose content (12.5%wmf) of the miscanthus H₁ fraction, in detriment to a reduction in the xylose (64.4%wmf) and acetyl group (16.5%) content.

Pine H₁ degradation rate profile was intermediate to those of the

previous H₁ fractions. It showed a maximum degradation rate around 280 °C, which is comparable to the behavior of deciduous wood H₁ fractions. However, a slight deceleration could be appreciated around 250 °C, inducing a small change in the degradation rate profile. Pine H₁ sugar composition mainly consists of mannose (49.2%wmf) and lower amounts of xylose (18.4% wmf) and glucose (14.8%wmf). This composition could explain that the maximum in the degradation pine H₁ is reached at higher temperatures, as mannose-based sugars were pointed out in the literature as less reactive than xylose-based ones [29].

Finally, the kinetic behavior of wheat straw H₁ was close to that of the other fractions until 250 °C, but with a slower acceleration. From this temperature, solid mass loss was significantly decelerated until 300 °C, by losing around 0.7%wmf·min⁻¹ in this period. The behavior of this fraction was checked as reproducible and could be justified by its high glucose content compared to the xylose content (Table 1). This might suggest that a high glucose content, rather typical of a cellulose-based fraction, would be susceptible to mitigate the degradation of a given hemicellulose fraction.

3.2.2.2. «Hemicelluloses 2» fraction (H₂). The final remaining mass loss, after the non-isothermal torrefaction step, was close for all the «hemicelluloses 2» fractions (H₂), between 24.9%wmf for miscanthus H₂ fraction and 33.3%wmf for wheat straw H₂ fraction (Fig. 6, left). Degradation rates were in the same order of magnitude as those of the corresponding H₁ fractions. They increased to around 3.1%wmf·min⁻¹ for ash-wood and pine H₂ fractions and to 4.7%wmf·min⁻¹ miscanthus H₂ fraction. In the end of the isothermal step, degradation rates decreased drastically, until values below 0.1%wmf·min⁻¹ at the end of this period (Fig. 6, right). By comparing both hemicellulose fractions per biomass, it was shown that extracted H₁ (Fig. 5) and H₂ fractions (Fig. 6) present a different behavior.

Regarding the remaining solid mass loss profiles (Fig. 6, left), ash-wood and pine H₂ fractions presented a slightly higher mass loss below 300 °C, while the degradation of wheat straw H₂ fraction was the lowest. This behavior might be due to the higher mannose content of this last fraction (33.3%, Table 1), while the other H₂ fractions were principally composed of xylose [29]. However, the low mannose content of pine (7.5%) and miscanthus (2%) H₂ fractions did not seem to impact their behavior in torrefaction, at least below 300 °C, as they presented different degradation profiles. All

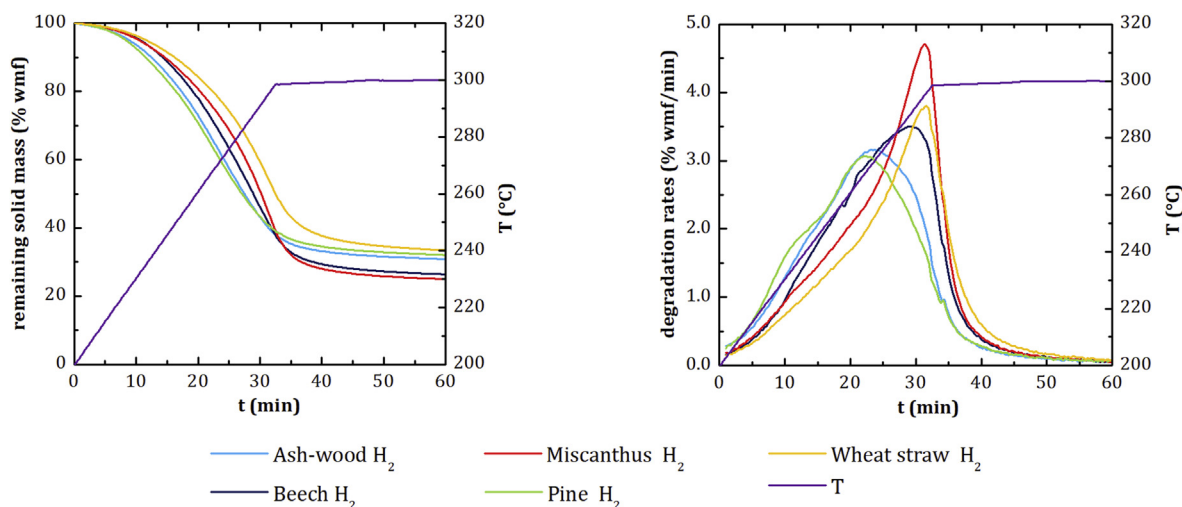


Fig. 6. Remaining solid mass (left) and degradation rates (right) versus time and temperature in torrefaction in TGA-GC/MS for the H₂ fractions from the biomasses of study.

H₂ fractions presented a low glucose content (6.2–16.7%), which might contribute to the stabilization of solid mass loss at 300 °C (Fig. 6).

A detailed analysis of degradation rate curves (Fig. 6, right) showed that ash-wood, beech and pine fractions presented a global acceleration of solid mass loss interrupted by a brief deceleration at around 240 °C, which is especially remarkable for pine H₂ fraction. After that, these three fractions reached the maximum degradation rate at around 275 °C. Wheat straw and miscanthus H₂ fractions presented an acceleration of the mass loss until the maximum degradation rate, attained around 295 °C. In both cases, the maximum of the degradation rate for each sample was attained at temperatures slightly higher than those for H₁ fractions (Fig. 5, right). These maxima were followed by a deceleration of the solid mass loss which tended to stabilize in the isothermal stage.

To sum up, sugar composition strongly impacted hemicelluloses behavior in torrefaction, both in terms of extent of the degradation and remaining solid mass profile in function of time and temperature. Even if biomass type influenced sugar proportions in woody and agricultural biomasses (i.e. glucomannans in coniferous wood), it was shown to have a lower impact on hemicelluloses degradation than neutral monosugars composition.

3.2.3. Lignin fraction (L)

The remaining solid mass at the end of the torrefaction experiment was few variable for all « lignin » extracted fractions (L), around 70.0 %wmf, except for pine L fraction (65.1 %wmf, Fig. 7, left). Degradation rates were low, below 1.1% wmf·min⁻¹, for all L fractions (Fig. 7, right). In this case, as L fractions could not be characterized in terms of H-, G- and S- unit content, lignin composition as described in the literature will be considered [13,25,68].

L fractions showed three solid kinetic patterns in the non-isothermal torrefaction step (0–33.3 min, Fig. 7, left), represented by the behaviors of deciduous wood L fractions (ash-wood and beech), agricultural by-products and herbaceous crops L fractions (wheat straw and miscanthus) and coniferous wood L fraction (pine). These patterns were confirmed by the degradation rates curves (Fig. 7, right). More precisely, ash-wood and beech L fractions presented a rather constant degradation rate acceleration until 300 °C, with a slight change in the degradation rate curve around 250 °C. This change was more pronounced for wheat straw and miscanthus L fractions, as their degradation rate was increasing from the beginning of the torrefaction until its stabilization around 250 °C. Pine L degradation rate increased until around 235 °C and

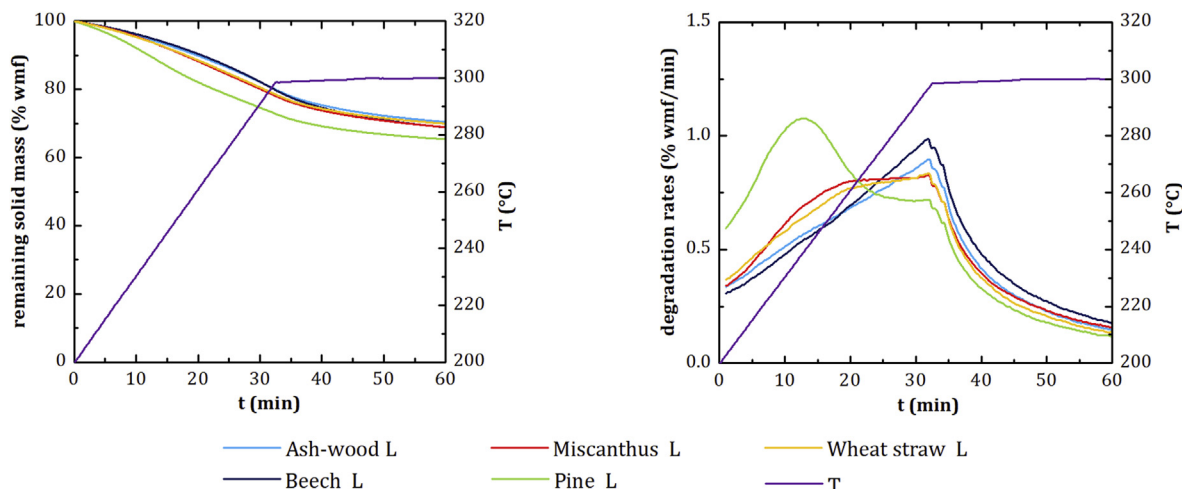


Fig. 7. Remaining solid mass (left) and degradation rates (right) versus time and temperature in torrefaction in TGA-GC/MS for the L fractions from the biomasses of study.

then decreased, to finally stabilize from 275 °C. By considering the isothermal torrefaction step, all L fractions presented a progressive decrease of their degradation rate. The final low solid mass loss measured for all lignin fractions, as well as the low degradation rates observed, are in agreement with the partial degradation of lignin in the torrefaction temperature range reported in the literature [8,17]. Furthermore, the differences in lignin structure according to biomass type might explain these observations: G-type lignin (mostly in pine lignin) would seem to be more easily degraded than H-type lignin (all biomasses), itself more easily degraded than S-type lignin (deciduous wood, mainly). However, these differences in the degradation patterns might also be due to cleavage of internal bonds in lignin units.

3.2.4. Evolution of the elemental composition of the extracted fractions

The elemental composition of the solid was characterized before and after torrefaction for C^{II} (Fig. 8A), DMSOr (Fig. 8B), H₁ (Fig. 8C) and L (Fig. 8D) fractions.

Small changes in elemental composition were observed for C^{II} (Fig. 8A), DMSOr (Fig. 8B) and L (Fig. 8D) fractions, except pine L

fraction, which was richer in carbon. In these cases, a similar result was shown for each extracted fraction from different biomasses, represented by a low dispersion of the points in the ternary diagrams. This behavior was in agreement with the small differences observed in the remaining solid mass profiles of the corresponding extracted fractions (Figs. 3–7). On the contrary, the evolution of H₁ fractions elemental composition was shown to be dependent on the nature of the hemicellulose fractions (Fig. 8C). Their transformation was very limited for wheat straw H₁ and more pronounced for the woody samples. However, remaining solid mass profiles were not directly correlated to changes in elemental composition of H₁ fractions. These results confirmed the influence of the sugar composition on hemicelluloses conversion through torrefaction.

The increase in carbon content in the torrefied C^{II} fractions was between 22.3 and 29.2%, while their oxygen content was reduced by 15.3–22.2%. In the case of H₁ fractions, the enrichment of the carbon content in the solid went from 10.5 to 50.1%, in detriment to a decrease in the oxygen content from 6.2 to 42.4% and sensible changes in the hydrogen content. It is noteworthy that the wheat straw sample suffered a more pronounced decrease of its hydrogen

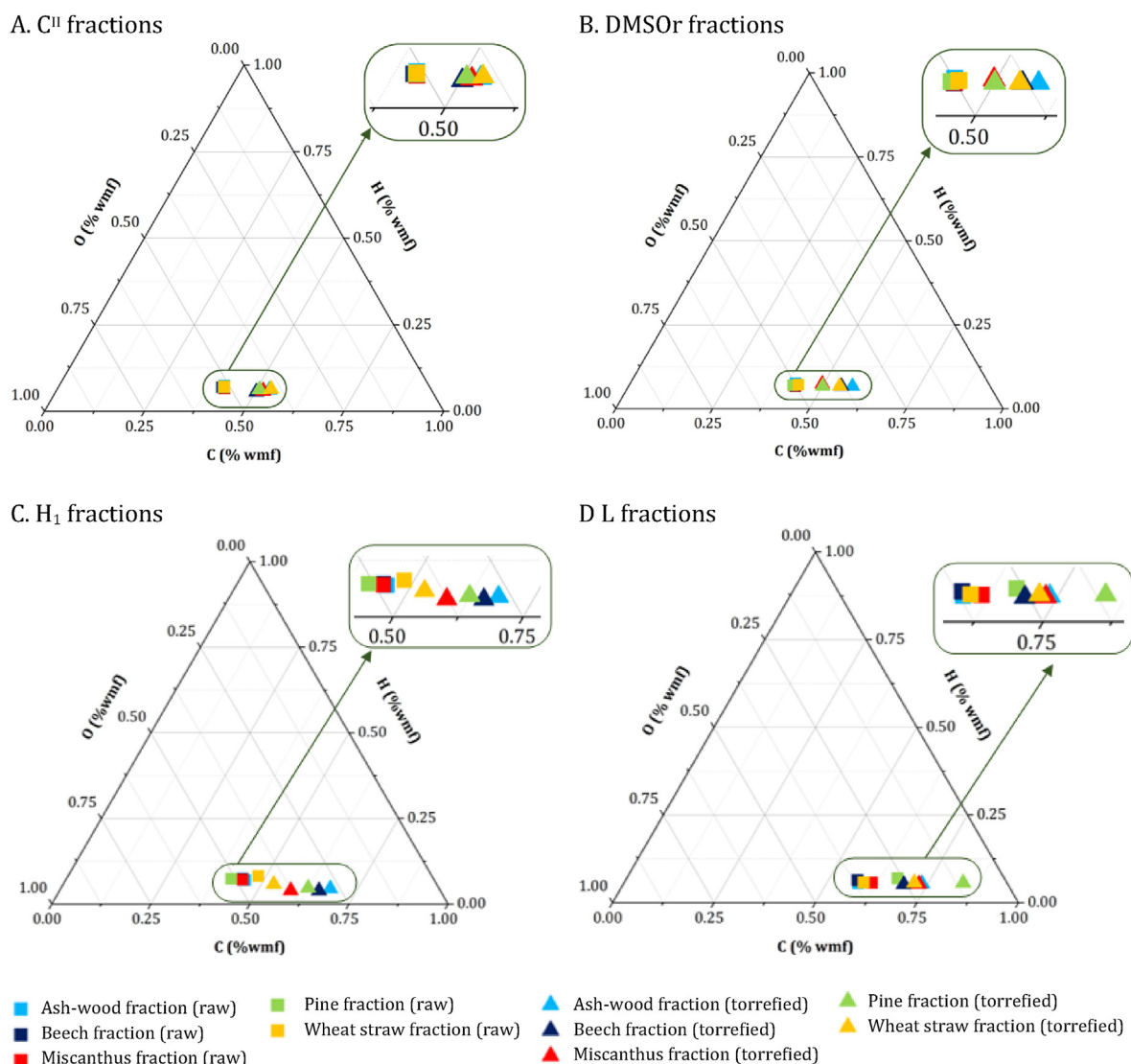


Fig. 8. Ternary diagram of CHO elemental composition of raw and torrefied C^{II} (8A), DMSOr (8B), H₁ (8C) and L (8D) fractions in TGA-GC/MS.

content compared to its oxygen content, in proportion (28.7 and 6.2%, respectively). The carbon content enrichment of the torrefied L fractions was between 19.2 and 27.3%, comparable to that of C^{II} fractions. This was associated with a decrease in oxygen from 29.6 to 59.1%. This diminution might be explained by a high production of permanent gases and/or oxygenated phenolic compounds (not measured), according to previous studies in the literature [13].

These results enlighten the fact that the C, H, O elemental composition does not seem to be sufficient to differentiate the extracted fractions during torrefaction.

4. Conclusions

An optimized extraction procedure was proposed to obtain cellulose, hemicellulose and lignin fractions from five woody and agricultural biomasses. The torrefaction of the obtained extracted fractions showed different solid degradation profiles from those previously reported in the literature for commercial compounds. This result highlighted the importance of deeply characterizing the chemical compounds selected as representative for biomass modelling. Furthermore, biomass family (deciduous wood, coniferous wood, agricultural biomasses and herbaceous crops) may strongly impact the behavior of macromolecular components in torrefaction, especially for hemicelluloses and lignin.

The five C^{II} fractions were few degraded in torrefaction. Their degradation started at lower temperatures (around 230 °C) than those previously reported in the literature for commercial micro-crystalline cellulose (around 300 °C). C^{II} fractions presented a lower level of hemicellulose-sugars impurities than that reported for commercial cellulose. However, their allomorphic structure was different from that of the native cellulose in biomass. The comparison of the behavior of the DMSO with that of the C^{II} fractions in torrefaction may suggest a minor impact of cellulose allomorphic structure, while the presence of hemicellulose sugars enhance the degradation of a cellulose-based fraction, especially below 300 °C.

The strong and fast degradation undergone by hemicelluloses in torrefaction was shown to be mainly conditioned by the sugar composition, especially by xylose and mannose. Furthermore, this degradation was different for the fractions extracted from the five biomasses of study. This behavior highlights the influence of the biomass type, which determines the proportions and the combination of sugars in hemicelluloses, on their degradation in torrefaction. As the behavior of H₁ and H₂ fractions for the same biomass was different due to their different sugar composition, both hemicellulose extracted fractions per biomass seem to be complementary to describe the global hemicellulose behavior in torrefaction.

L fractions degradation was shown to be very dependent on its composition in H- (phenyl-), G- (guaiacyl-) and S- (syringyl-) units. According to the results, three extracted lignin fractions would be enough to represent the lignin degradation pattern in biomass torrefaction, namely pine lignin (principally G- units), a deciduous wood lignin (G- and S-units) and an agricultural lignin (G-, S-, H-units).

The evolution of the solid elemental composition through torrefaction, expressed by its major composition in carbon, oxygen and hydrogen, was only notable for H₁ fractions. Lignin and cellulose present only moderate changes in their composition, due to their limited degradation at torrefaction temperatures. As a result, considering the main elemental composition instead of biomass macromolecular components seem to be insufficient, while biomass main macromolecular components would lead to a more accurate description of biomass behavior in torrefaction.

CRedit authorship contribution statement

María González Martínez: Conceptualization, Investigation, Formal analysis, Methodology, Visualization, Writing - original draft. **Capucine Dupont:** Conceptualization, Supervision, Project administration, Funding acquisition. **Denilson da Silva Perez:** Conceptualization, Methodology, Resources, Investigation. **Gérard Mortha:** Resources, Investigation. **Sébastien Thiéry:** Investigation. **Xuàn-mi Meyer:** Methodology, Supervision, Validation. **Christophe Gourdon:** Methodology, Supervision, Validation.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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